

Fertility Preservation In Reproductive Age Woman Facing Gonadotoxic Treatments

Jeff Roberts, MD, FRCSC Niamh Tallon, MB BCh BAO, FRCSC Hananel Holzer, MD

CFAS Clinical Practice Guideline Committee

William Buckett, MD Jon Havelock, MD Hananel Holzer, MD Neal Mahutte, MD Jason Min, MD (Chair) Jeff Roberts, MD Sony Sierra, MD Camille Sylvestre, MD Beth Taylor, MD

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Reproductive challenges for young women undergoing gonadotoxic treatments

Modern cancer treatments for young women have improved cure rates, but more often than not the price paid for survival is the loss of reproductive function from gonadal toxicity. High-dose alkylating agents and ionizing radiation have well recognized gonadotoxicity, inducing sterility in a high proportion of patients. Breast cancer affects more than 24,000 Canadian women per year. Fifteen per cent of these women are of reproductive age, making it the most common malignancy in this age group and the representing the bulk of referrals to assisted reproductive technology (ART) facilities for fertility preservation¹. Other cancers seen in young women include hematologic (lymphoma and leukemia), endometrial, and cervical. Less commonly, patients with autoimmune disorders like systemic lupus erythematosus (SLE) and hematologic conditions will require chemotherapeutic agents for medical management. Gonadal function and fertility outcomes have improved greatly with the newer regimens, however patients and physicians alike need to be aware of the deleterious effects that these treatments have on reproduction.

Management of the young woman with cancer presents several unique medical and social challenges. In general these patients

are ill-prepared for the diagnosis of cancer and the reality of their own mortality. With a limited understanding of assisted reproductive technologies or even basic fertility issues, patients are unlikely to seek medical advice on fertility preservation. Breast cancer patients under 40 years of age are also confronted with more aggressive tumor grades and reduced disease-free survival²⁻⁴. Combination chemotherapy regimens for breast cancer are delivering ever-increasing 5 year survival rates for early-stage disease, and further reductions in mortality are achieved with the addition of adjuvant hormonal therapies for patients with estrogen receptor positive tumors⁵. However, given the improved survival with all tumor types and stages, the need for adequate fertility counseling and a multidisciplinary team approach for these patients have never been greater⁶. Patients facing potentially sterilizing chemotherapy and radiotherapy can now benefit from recent advances in cryopreservation techniques which allow for the banking of oocytes, embryos, and ovarian tissue without compromising survival. With this CFAS guideline we outline the current understanding of the pathophysiology of gonadotoxicity from cancer treatments, the methods to minimize this damage, and the use of medical and surgical reproductive technologies to allow for future fertility and pregnancy. Evidence is graded as outlined in the report of the Canadian Task Force on Preventative Health Care (Table1).^{7,8}

Table 1: Quality of evidence assessment and classification of recommendations as defined by the Canadian Task Force on Preventative Health Care

Quality of Evidence Assessment ⁷	Classification of Recommendations ⁸
I: Evidence obtained from at least one properly randomized controlled trial	A. There is good evidence to recommend the clinical preventive action
II-1: Evidence from well-designed controlled tri-	B. There is fair evidence to recommend the clini-
als without randomization	cal preventive action
II-2 Evidence from well-designed cohort (pro-	C. The existing evidence is conflicting and does
spective or retrospective) or case-control stud-	not allow a recommendation for or against use
ies, preferably from more than one centre or	of the clinical preventive action; however, other
research group	factors may influence decision-making



Quality of Evidence Assessment ⁷	Classification of Recommendations ⁸
II-3 Evidence obtained from comparisons be- tween times or places with or without the inter- vention. Dramatic results in uncontrolled ex- periments (such as the results of treatment with penicillin in the 1940s) could also be included in the category	D. There is fair evidence to recommend against the clinical preventive action
III Opinions of respected authorities, based on clinical experience, descriptive studies, or reports of expert committees	E. There is good evidence to recommend against the clinical preventive action
	L. There is insufficient evidence (in quantity or quality) to make a recommendation; however, other factors may influence decision-making

Chemotherapy

Ovarian reserve represents the population of primordial follicles within the ovary, which make up more than 90% of the follicular population at any given time. The ovarian pool of oocytes and their individual reproductive potential declines with age, as reflected in diminishing fecundity rates and pregnancy rates with medical fertility treatments⁹⁻¹¹. Chemotherapeutic agents appear simply to accelerate this process¹². Gonadotoxicity of combination chemotherapeutic treatments vary according to the specific agents used, cumulative doses, protocol, and reproductive potential of the patient at the time of treatment^{13,14}. Cyclophosphamide and other alkylating agents are the most toxic to the ovary, inflicting a dose-dependant exponential decline in primordial follicle density^{15, 16}. Compared to other regimens, cyclophosphamide-containing protocols are four times more likely to result in ovarian failure, with almost 80% of cases occurring within the first year¹³.

Protocols are classified into low, intermediate, or high risk of inducing ovarian failure, with the incidence of menopause ranging from less than 20% to over 80%^{17, 18}.

Quantifying the gonadotoxic effects of each chemotherapeutic regimen is difficult and poorly studied to date. Most existing clinical trials and population studies for chemotherapeutic agents report the incidence of premature ovarian failure (POF) and ovulatory dysfunction as the measure of fertility. Infertility and diminished ovarian reserve are typically associated with eumenorrhea and ovulatory cycles¹⁹. Destruction of pre-ovulatory follicles results in temporary amenorrhea for a period of 3 to 4 months however long-term ovarian function can be maintained by as little as 10% of the ovary, so clinical measures of menstrual function are a poor assessment of ovarian damage²⁰. In the absence of long term follow-up of fertility and pregnancy outcomes, the effects of cancer treatment on future reproductive function will be underestimated.

As expected, the incidence of acute ovarian failure, infertility and early menopause in chemotherapy patients correlates with age¹⁹. Regardless of the type of chemotherapeutic agents administered, at least a fraction of ovarian reserve will be lost even if this is not immediately apparent with clinical and laboratory evaluation. Most objective measures of ovarian reserve are altered by chemotherapy²¹. Low risk treatments like ABVD (adriamycin, bleomycin, vinblastine

and dacarbazine) for Hodgkins lymphoma appear to have minimal short-term effects on reproduction in women under 30, however clear effects are seen in both menstrual function and ovarian reserve testing in older age groups²². Even if a patient is deemed to be at "low risk" for premature menopause, a shorter reproductive life can be expected even if regular menstrual cycles resume^{23, 24}. Anecdotal experience would suggest that cancer survivors with a history of chemotherapy have poor outcomes with medical fertility treatments. Youth is certainly a protective factor, but long term follow-up of childhood cancer patients demonstrate clear effects on ovarian reserve and reproductive potential later in life²⁵⁻³¹. Fortunately chemotherapeutic agents do not appear to have long-term effects on the genetic competency of surviving oocytes, or the future pregnancies themselves³², but based on murine data the risk of fetal malformation may by elevated for up to 6 to 12 months after exposure³³. Overall, the most commonly used combination chemotherapies likely advance a woman's reproductive age by 10 years, with the onset of menopause dependant on the patient's ovarian reserve at the start of treatment³⁴.

Radiation therapy

As with chemotherapeutic agents, the impact of ionizing radiation on the female reproductive tract relates to the age at exposure and effective dose (fractionation schedule)^{35, 36}. Abdominopelvic irradiation can lead to high rates of premature ovarian failure, with even less than 2 Gy causing loss of over 50% of the primordial follicle pool (LD50)^{37, 38}. By comparison, typical doses for gynecologic malignancies total 50 Gy. Hypothalamic-pituitary-gonadal function can be impaired by cranial irradiation, with the highest incidence of central hypogonadism occurring with doses above 30-40 Gy^{39, 40}. Other risk factors for POF include concurrent alkylating agent administration, high dose radiation and the diagnosis of Hodgkins lymphoma³⁸.

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Pelvic radiation impairs fertility and is associated with poor pregnancy outcomes, including early and midtrimester loss, preterm birth and low birthweight⁴¹. The pathophysiology appears to involve vascular, endometrial and myometrial damage^{31, 42}. Exposure prior to completion of puberty impairs normal uterine development, with a resulting reduced adult uterine volume which is refractory to estrogen replacement therapy. The potential need for gestational surrogacy should be discussed in these cases.

Early access to care and barriers to referral

In 2006, the American Society of Clinical Oncology (ASCO) set out to provide guidance to oncologists regarding fertility preservation and concluded that the process of informed consent requires a discussion of future fertility issues and options for fertility preservation¹⁷. An algorithm was suggested, which included provision of this counselling from a structured group which includes the medical oncologist, reproductive endocrinologist and psychologist. Ideally, such a collaborative multidisciplinary team would satisfy the need for a patient-centered approach to determining a realistic likelihood of success, given all the factors that can play into such a multifaceted issue. Recently, a collaborative group was established in Canada called the Cancer Knowledge Network which endeavors to educate patients and professionals, and to connect patients with their regional fertility preservation services (www.cancerkn.com).

Despite the ASCO recommendation, some cancer specialists do not routinely discuss fertility preservation, with nearly half never referring to a fertility specialist⁴³. Many barriers have been identified, including a lack of knowledge of fertility preservation options⁴⁴ and available local resources^{45,46}, and the perception that assisted reproductive technologies are cost prohibitive and of limited efficacy^{47, 48}.



The constraints of time and concern for cancer treatment delay was also cited, however early involvement of the fertility specialist is critical for the provision of timely fertility preservation services^{49, 50}. Directly comparing patients undergoing fertility preservation treatments to those with standard cancer treatment protocols, there were no statistical or clinical differences between the groups regarding time from initial diagnosis to chemotherapy initiation or in the time from definitive operation to chemotherapy $(p = < 0.27 \text{ and } p = < 0.79 \text{ respectively})^{51}$. The median time from referral to oocyte retrieval was 32 days (range 13-66 days)⁵¹ with a multidisciplinary team functioning to provide IVF within 18 days from referral (median of 11 days for ovarian stimulation)⁵². Chemotherapy may actually start up to 3 weeks earlier if a fertility preservation referral is made before cancer surgery⁵³. Collaborative efforts between the fertility specialist and oncology team should aim to provide informed counseling regarding future infertility and the suitability of individualized fertility preservation treatments⁵⁴. This requires early referral and timely consultation with a fertility specialist, with the provision of fertility preservation treatment in conjunction with the oncologic management schedule.

Recommendations

After the diagnosis of cancer or other medical conditions requiring potentially sterilizing medical or surgical treatments in a reproductiveage woman, an immediate referral to a reproductive endocrinologists and infertility (REI) specialist is strongly suggested, to provide patients with counseling regarding their fertility and fertility preservation management options. (II-B)

A multidisciplinary network to facilitate referrals to professionals with expertise in fertility preservation should be considered. (II-3B)

Assessment of the young cancer patient

Prior to considering fertility preservation treatments the individualized risk that the cancer treatment poses to the patient's future fertility must be considered. In the case of breast cancer we should be mindful of the two year period of observation following completion of chemotherapy, and lengthy delays when adjuvant hormone therapies are employed. It is important that the oncology team is consulted prior to initiating fertility preservation treatment. Careful coordination of the fertility preservation treatments may be required to allow for timely delivery of the cancer treatment, with a clear understanding with the patient that her cure takes precedence over fertility.

Ovarian reserve testing should be considered to aid in the developing the ovarian stimulation protocol and to provide a reasonable estimate of her age-related prognosis. Basal follicle stimulating hormone (FSH) has been the standard evaluation of ovarian reserve for many years and a simple means to screen patients for diminished response and poor outcome from IVF⁵⁵. Antimullerian hormone (AMH) is proving to be the most predictive for ovarian response to exogenous gonadotropins and pregnancy outcome and also the most versatile in these patients⁵⁶⁻⁵⁸. AMH is detectable at all ages, and unlike FSH, is stable throughout the menstrual cycle so can be performed at the time of presentation. In combination with the patient's age, AMH can help assess future fertility through the quanitfying of the short-term chemotoxic effects on ovarian reserve⁵⁹, providing an estimated age of menopause^{60,61}, and assessing the patient's susceptibility to the gonadotoxic effects of chemotherapy^{62,63}. Similar to FSH, AMH is a better predictor of oocyte numbers and ovarian response to gonadotropins, than it is of successful pregnancy^{64, 65}. A complete transvaginal pelvic ultrasound with antral follicle count (AFC) is an essential part of the basic fertility



assessment of these patients⁶⁶. In addition to providing further data about the patient's ovarian reserve^{67, 68}, pre-treatment ultrasound also evaluates for pelvic pathology and adnexal anatomy in preparation for controlled ovarian stimulation and oocyte harvest.

Recommendations

Serum follicle stimulating hormone (FSH), antimullerian hormone (AMH) levels, and/or antral follicle count (AFC) should be performed prior to chemotherapy to assist in the selection of gonadotropin doses and to prognosticate gonadotoxic effects of chemotherapy. (II-2B)

Follow-up serum FSH and AMH should be considered for assessing the gonadotoxic effects of chemotherapy. (II-2A)

Fertility preservation options

On the basis of a fertility assessment, oncologic treatment plan, and the patient's reproductive needs, individualized fertility preservation plans can be formulated formost patients. The decision to proceed with fertility preservation treatments should take into account age, diagnosis, oncology treatment regimen, reproductive potential with and without treatment, and the patient's personal/social situation. More universally, gonadotropin releasing hormone (GnRH) agonists are administered concurrently with cytotoxic treatments in an effort to provide some level of protection and are suitable for all ages. Assisted reproductive technologies have been used to generate oocytes and embryos for cryopreservation and future use. Creation of embryos requires sperm from a partner, or when there is no partner the use of donor sperm. Oocyte vitrification is proving to be an excellent option for women even when a partner is present because it provides the patient with reproductive autonomy. In vitro maturation

(IVM) is an investigational strategy available through a limited number of centers. Whilst also investigational, ovarian tissue cryopreservation serves as the most hopeful option for children and young adolescents who are otherwise limited by their reproductive immaturity.

Gonadotropin-releasing hormone (GnRH) agonists

Reports of reduced amenorrhea rates in young women using adjuvant gonadotropinreleasing hormone agonists (GnRHa) prompted the investigation of their chemoprotective properties in the ovary. Despite the limited evidence for their efficacy, these agents are currently used routinely by some centres during chemotherapy. Proposed mechanisms of action include hypogonadotropism-induced ovarian quiescence, reduction of ovarian blood flow and agonistic effects on ovarian GnRH receptors. Three small prospective randomized studies have addressed this question with two demonstrating a reduction in premature ovarian failure, one from 66.6% to 11.4%⁶⁹⁻⁷¹. A meta-analysis also showed a protective effect, however most studies have used historical controls⁷². A potential concern with the use of GnRH agonists in breast cancer patients is that the resulting hypoestrogenic state may inadvertently arrest malignant cells in the resting (G0) phase, and render them less susceptible to chemotherapy^{73, 74}. Larger studies are required to better evaluate the efficacy of these agents.

Recommendation

GnRH agonists can be considered as a means of gonadal cytoprotection prior to combination chemotherapy. (I-B)



Embryo cryopreservation

Cryopreservation of embryos is a standard technique employed by all IVF clinics for the banking of supernumerary embryos and for situations in which the transfer of fresh embryos is ill-advised, such as with severe cases of ovarian hyperstimulation syndrome⁷⁵.

As a method of fertility preservation, it has been available to cancer patients for many years. Foremost is the need for a male partner unless the patient is prepared to use donor sperm. The patient's chance of a successful future pregnancy depends on the number of high quality embryos obtained; in turn the number of IVF cycles performed, and the time available to achieve adequate stimulation. In 2012, the clinical pregnancy rate for frozen embryo transfer among the 33 IVF facilities in Canada was 34%⁷⁶. With all of the necessary resources at hand, many IVF facilities routinely cryopreserve embryos as a means of fertility preservation in patients with male partners or those that wish to use donor sperm.

Recommendation

Embryo cryopreservation is a recommended method of fertility preservation. (I-A)

Oocyte cryopreservation

For women without a male partner or women desiring reproductive autonomy, oocyte cryopreservation has become the standard approach. Historically, the technique has been beset by lower pregnancy rates compared to embryo cryopreservation, but with recent advances in cryotechnology the service is becoming more widely available⁷⁷. The low pregnancy rates were related to several technical challenges encountered during the freeze-thaw process and the in vitro maturation

of immature oocytes. Mature oocytes provide the best chance for pregnancy, but have several characteristics that make them susceptible to cryodamage. The oocyte's large size (low surface area to volume ratio) and high water content also makes it vulnerable to ice crystal formation, rupture, and limited penetration of cryoprotectant solutions⁷⁸. Since mature oocytes are arrested in metaphase II, the spindle apparatus is fully extended and prone to disassembly at lower temperature, with subsequent chromosome dispersion and aneuploidy^{79,80}. Despite the potential obstacles, clinical and neonatal outcomes to date attest to the safety of this technology⁸¹. The efficiency, feasibility and safety of the technology have developed to the point that the American Society for Reproductive Medicine (ASRM) no long considers it experimental for the purpose of fertility preservation in women undergoing gonadotoxic therapies^{82, 83}.

Vitrificationhasbeenintegral to the improvements and success of oocyte cryopreservation. The technique directly solidifies the oocyte and surrounding solution into a glasslike (vitreous) state, minimizing the formation of potentially disruptive intracellular and extracellular ice crystals. Meta-analyses support the superior thawing survival and clinical outcomes of oocyte vitrification⁸⁴⁻⁸⁶.

randomized controlled А recent trial demonstrated clinical equivalency of vitrified oocytes to fresh in the setting of anonymous oocyte donation⁸⁷. Other groups are also starting to report similar success and the technique has quickly become the standard approach for both oocyte and embryo cryopreservation⁸⁸. With refinements in technique and better clinical outcomes, oocyte cryopreservation is proving to be a simple and versatile method of fertility preservation that will provide women with reproductive autonomy.



Recommendation

Oocyte cryopreservation by vitrification is a recommended method of fertility preservation. (I-A)

Controlled ovarian stimulation of the cancer patients

By employing modern IVF protocols and appropriate doses of gonadotropins, oocyte and embryo yield and ultimately the number of future attempts for pregnancy can be maximized. In the case of breast cancer, we are commonly restricted to a period of four to six weeks between the time of surgery and chemotherapy, only allowing for one or two IVF cycle attempts since gonadotropins are traditionally initiated with menses.

GnRH antagonist protocols provide the most flexibility during ovarian stimulation. GnRH antagonist treatments are shorter, require less gonadotropin, and reduce the risk of ovarian hyperstimulation syndrome (OHSS)⁸⁹⁻ Gonadotropins can be initiated with spontaneous menses, through truncation of the menstrual cycle with the administration of a GnRH antagonist shortly following ovulation⁹³, or randomly throughout the patient cycle⁹⁴⁻⁹⁸. The dose of gonadotropins should be individualized to the patient based on age and ovarian reserve testing, with the goal of maximizing the number of high grade embryos at the end of the process, without compromising the patient's medical status prior to starting her cancer Minimal data exists to suggest treatments. that these patients have different gonadotropin requirements or that the quality of the oocytes and embryos are compromised by their illness, however dosing decisions should be left to a physician with experience using gonadotropins in these patients⁹⁹. In an effort to minimize the patient's estrogen exposure after oocyte retrieval and her risk of early OHSS, GnRH agonists can be employed with antagonist cycles for triggering final oocyte maturation¹⁰⁰⁻¹⁰². Similar to oocyte donors, these patients are not at risk for the more clinically worrisome late OHSS since they are not conceiving. Since an inadequate induction of the luteinizing hormone (LH) surge is the principle risk of this technique, it is also acceptable to reserve GnRH agonist "trigger" for patients with a hyperresponse, or to supplement with a small dose of hCG^{101, 103, 104}. GnRH antagonist protocols are preferred in these cases for several reasons but are often the most practical option given that gonadotropins can be started quickly.

Long GnRH agonist protocols are still used for controlled ovarian stimulation technique but have the well-recognized risk of inducing a luteal "flare" from the pituitary with rescue of the corpus luteum and functional ovarian cysts, leading to treatment delays¹⁰⁵.

Recommendations

Ovarian stimulation protocols utilizing GnRH antagonists should be considered for embryo and oocyte cryopreservation. (II-3B)

GnRH agonists are recommended for the induction of oocyte maturation when utilizing GnRH antagonist cycles in an effort to minimize the risk of ovarian hyperstimulation syndrome. (I-2A)

Use of cytoprotective agents during ovarian stimulation in breast cancer patients

Many breast cancer tumor cells are estrogen receptor positive, and accordingly are susceptible to environments with estrogen excess^{106, 107}. Even tumors that are classified as receptor negative will contain a small percentage of receptor-positive cells^{108, 109}.



Serum estradiol levels reach supraphysiologic levels during controlled ovarian stimulation, typically 5000 pmol/mL (peak natural cycle levels 750 to 1300 pmol/mL) and not uncommonly exceeding 10,000 pmol/mL. Although no clinical data currently exist, high levels of estrogens could in theory stimulate subclinical disseminated disease¹¹⁰, so any therapy that antagonizes this effect is reasonable¹¹¹⁻¹¹³.

Two strategies are commonly employed to minimize this estrogen exposure: recovering oocytes from an unstimulated IVF cycle, or administering cytoprotective agents in combination with gonadotropin stimulation. Aromatase inhibitors have proven efficacious as an adjuvant therapy for the management of micrometastatic disease^{111, 114-116}. Thev have gained popularity as ovulation induction agents and adjuncts in IVF protocols¹¹⁷. More importantly for the breast cancer patient, they suppress estradiol production during IVF stimulation. Letrozole is the most potent of the aromatase inhibitors, suppressing greater than 96% of its activity. With concurrent use of aromatase inhibitors, gonadotropins can be administered to maximize embryo yield while minimizing estradiol levels^{118, 119}. Ultrasound follicle tracking serves as the only measure for dosing adjustment and assessing the patient's risk for OHSS. Since the ovary remains hyperstimulated well beyond retrieval of the oocytes, sustained use of letrozole for at least another seven days is recommended.

Recommendation

In women with breast cancer and other estrogensensitive diseases, aromatase inhibitors should be considered when administering gonadotropins. (II-3B)

Ovarian cryopreservation and transplantation

Any patient receiving chemotherapy or radiotherapy that targets the ovarian follicles can be considered a candidate for ovarian cryopreservation, assuming that she is at low risk for ovarian metastasis¹²⁰. Some patients may also not have sufficient time to undergo ovarian stimulation for oocyte or embryo freezing, or have abdominal surgery as part of their oncology plan.

Since the first experiments with ovarian transplantation in animals¹²¹, steady advances have been made in humans. Since then, other species have been successfully transplanted with autologous ovarian tissue, as well as with human xenografts¹²². With the knowledge gained from these experiments, trials in human transplantation were initiated. In the case of cancer patients, caution should be exercised to prevent the reintroduction of disease. Transmission of cancer has been demonstrated in the animal model^{123, 124}, and metastatic seeding of the ovary occurs commonly with some cancers. In the case of BRCA, the risk of ovarian cancer is significant so screening for this mutation should be considered in all breast cancer patients before ovarian tissue harvest¹²⁵.

At an appropriate time after completion of the patient's cancer therapy, the tissue is thawed and transplanted either orthotopically or heterotopically within subcutaneous tissue or pelvis¹²⁶⁻¹³⁰. The major barrier for this technology is the delayed revascularization and resulting ischemia, fibrosis and subsequent loss of the primordial follicles^{121, 131}. Grafts become hormonally active three to four months after transplantation, at which time oocyte harvesting can be attempted, with or without the aid of exogenous gonadotropins to stimulate follicle development. To date over 23 live births have been reported in humans, however some may have originated from the contralateral



in situ ovary since only one ovary is typically harvested¹³²⁻¹³⁸. Several groups are presently experimenting with whole ovary vitrification and transplantation as a means to improve efficiency and clinical outcomes¹³⁹⁻¹⁴². With the limited success of this technology to date, the potential for reseeding of metastatic disease and surgical risks, ovarian transplantation should still be considered investigational and limited to cases where oophorectomy is planned. The procedure is further limited by the limited number of individuals and facilities with expertise in this technique.

Recommendation

Ovarian tissue cryopreservation and transplantation is investigational and should be limited to cases of oophorectomy or other predetermined abdominal surgeries by surgeons with the necessary experience, and under a clinical research approved by a research ethics board. (II-3B)

Pregnancy after cancer

There is no consensus on the best time to conceive after cancer treatments. Since most recurrences occur in the first two years patients are commonly asked by their oncologist to wait. However, some studies suggest that early conception does not negatively impact survival^{143, 144}. Five year survival is actually higher in breast cancer patients who achieve a pregnancy^{145, 146}, although this observation may certainly be accounted for by a healthy patient bias. Ultimately, the decision on timing of pregnancy should be determined in consultation with a cancer specialist.

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Fertility sparing surgical options

The harmful effects of radiation on the ovaries can be minimized by ovarian transposition. The ovaries are surgically transposed to a location outside the radiation field¹⁴⁷. This may also be combined with gonadal shielding to reduce the dose effect further¹⁴⁸. Using this technique, reports on the preservation of menstrual function have ranged from 65% to over 88%¹⁴⁹⁻¹⁵¹. The risks associated with the procedure include: ovarian cysts, adhesions, pelvic pain, ovarian migration, premature ovarian failure, and tubal injury^{20, 152}. A small risk of metastatic disease to the ovary exists in some malignancies, so transposition could facilitate the spread of disease^{20,153, 154}. Any benefits of transposition may be lost when adjuvant chemotherapy is employed^{155, 156}. Other fertility-sparing surgical options include cervical conization or trachelectomy for select early stage cervical cancer patients and unilateral oophorectomy or cystectomy in select ovarian neoplasms^{157, 158}.

Recommendation

Fertility sparing surgery should be considered when possible if it does not compromise survival. (III-B)

Ethical considerations

Standard treatments confer an overall net benefit to the patient, but when experimental the benefit must be carefully considered. This evolving field of oncofertility is aimed at improving outcomes from investigational procedures. For such interventions, the guidance of an institutional review board is recommended, with formal protocols and associated consent forms clearly stating the proposed treatment as investigational.



The consent process for women of all ages should include a discussion of the following:

- 1. Risk of cancer treatments on future fertility
- 2. Fertility preservation treatment options
- 3. Risks of delaying cancer treatment
- 4. Any investigational aspects of fertility preservation treatment
- 5. Realistic likelihood of success of the fertility preservation options
- 6. Potential risks of the fertility preservation treatment
- 7. Treatment costs
- 8. Disposition of human reproductive material
- 9. Posthumous reproduction
- 10. Alternative fertility options (oocyte donation, adoption, gestational surrogacy)

Summary

Recent strides in oocyte and embryo cryopreservation technology provide effective fertility preservation options through assisted reproductive technologies in Canada. A multidisciplinary approach and education of oncology professionals will help ensure that cancer patients receive the appropriate fertility preservation counselling and services.

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Bibliography

- 1. Canadian Cancer Society's Advisory Committee on Cancer Statistics. Canadian Cancer Statistics 2013. 2013.
- 2. Aebi S, Gelber S, Castiglione-Gertsch M, et al. Is chemotherapy alone adequate for young women with oestrogen-receptor-positive breast cancer? Lancet 2000; 355(9218): 1869-74.
- 3. Bentzon N, During M, Rasmussen BB, Mouridsen H, Kroman N. Prognostic effect of estrogen receptor status across age in primary breast cancer. International journal of cancer Journal international du cancer 2008; 122(5): 1089-94.
- 4. Adami HO, Malker B, Holmberg L, Persson I, Stone B. The relation between survival and age at diagnosis in breast cancer. The New England journal of medicine 1986; 315(9): 559-63.
- 5. Burstein HJ, Prestrud AA, Seidenfeld J, et al. American Society of Clinical Oncology clinical practice guideline: update on adjuvant endocrine therapy for women with hormone receptor-positive breast cancer. Journal of clinical oncology : official journal of the American Society of Clinical Oncology 2010; 28(23): 3784-96.
- 6. Jemal A, Thun MJ, Ries LA, et al. Annual report to the nation on the status of cancer, 1975-2005, featuring trends in lung cancer, tobacco use, and tobacco control. Journal of the National Cancer Institute 2008; 100(23): 1672-94.
- 7. Woolf SH, Battsta RN, Angerson GM, Logan AG, Eel W. Canadian Task Force on the Periodic Health Exam. Ottawa: Canada Communications Group p.xxxvii, 1994.
- 8. New grades for recommendations from the Canadian Task Force on Preventative Health Care. CMAJ 169:207, 2003.
- 9. Gunby J, Bissonnette F, Librach C, Cowan L, Fertility IVFDGotC, Andrology S. Assisted reproductive technologies (ART) in Canada: 2007 results from the Canadian ART Register. Fertility and sterility 2011; 95(2): 542-7 e1-10.
- 10. Tietze C. Reproductive span and rate of reproduction among Hutterite women. Fertility and sterility 1957; 8(1): 89-97.
- 11. van Noord-Zaadstra BM, Looman CW, Alsbach H, Habbema JD, te Velde ER, Karbaat J. Delaying childbearing: effect of age on fecundity and outcome of pregnancy. Bmj 1991; 302(6789): 1361-5.
- 12. Wallace WH, Kelsey TW. Human ovarian reserve from conception to the menopause. PloS one 2010; 5(1): e8772.
- 13. Goodwin PJ, Ennis M, Pritchard KI, Trudeau M, Hood N. Risk of menopause during the first year after breast cancer diagnosis. Journal of clinical oncology : official journal of the American Society of Clinical Oncology 1999; 17(8): 2365-70.
- 14. Arnon J, Meirow D, Lewis-Roness H, Ornoy A. Genetic and teratogenic effects of cancer treatments on gametes and embryos. Human reproduction update 2001; 7(4): 394-403.
- 15. Meirow D, Lewis H, Nugent D, Epstein M. Subclinical depletion of primordial follicular reserve in mice treated with cyclophosphamide: clinical importance and proposed accurate investigative tool. Human reproduction 1999; 14(7): 1903-7.
- 16. Walshe JM, Denduluri N, Swain SM. Amenorrhea in premenopausal women after adjuvant chemotherapy for breast cancer. Journal of clinical oncology : official journal of the American Society of Clinical Oncology 2006; 24(36): 5769-79.
- 17. Lee SJ, Schover LR, Partridge AH, et al. American Society of Clinical Oncology recommendations on fertility preservation in cancer patients. Journal of clinical oncology : official journal of the American Society of Clinical Oncology 2006; 24(18): 2917-31.
- 18. Hickey M, Peate M, Saunders CM, Friedlander M. Breast cancer in young women and its impact on reproductive function. Human reproduction update 2009; 15(3): 323-39.
- 19. Letourneau JM, Ebbel EE, Katz PP, et al. Acute ovarian failure underestimates age-specific reproductive impairment for young women undergoing chemotherapy for cancer. Cancer 2012; 118(7): 1933-9.
- 20. Wo JY, Viswanathan AN. Impact of radiotherapy on fertility, pregnancy, and neonatal outcomes in female cancer patients. International journal of radiation oncology, biology, physics 2009; 73(5): 1304-12.



- 21. Anderson RA, Themmen AP, Al-Qahtani A, Groome NP, Cameron DA. The effects of chemotherapy and long-term gonadotrophin suppression on the ovarian reserve in premenopausal women with breast cancer. Human reproduction 2006; 21(10): 2583-92.
- 22. Behringer K, Mueller H, Goergen H, et al. Gonadal function and fertility in survivors after Hodgkin lymphoma treatment within the German Hodgkin Study Group HD13 to HD15 trials. Journal of clinical oncology : official journal of the American Society of Clinical Oncology 2013; 31(2): 231-9.
- 23. Partridge A, Gelber S, Gelber RD, Castiglione-Gertsch M, Goldhirsch A, Winer E. Age of menopause among women who remain premenopausal following treatment for early breast cancer: long-term results from International Breast Cancer Study Group Trials V and VI. European journal of cancer 2007; 43(11): 1646-53.
- 24. Sklar C. Maintenance of ovarian function and risk of premature menopause related to cancer treatment. Journal of the National Cancer Institute Monographs 2005; (34): 25-7.
- 25. Larsen EC, Muller J, Schmiegelow K, Rechnitzer C, Andersen AN. Reduced ovarian function in long-term survivors of radiation- and chemotherapy-treated childhood cancer. The Journal of clinical endocrinology and metabolism 2003; 88(11): 5307-14.
- 26. Larsen EC, Muller J, Rechnitzer C, Schmiegelow K, Andersen AN. Diminished ovarian reserve in female childhood cancer survivors with regular menstrual cycles and basal FSH <10 IU/I. Human reproduction 2003; 18(2): 417-22.
- 27. Bath LE, Wallace WH, Shaw MP, Fitzpatrick C, Anderson RA. Depletion of ovarian reserve in young women after treatment for cancer in childhood: detection by anti-Mullerian hormone, inhibin B and ovarian ultrasound. Human reproduction 2003; 18(11): 2368-74.
- 28. Johnston RJ, Wallace WH. Normal ovarian function and assessment of ovarian reserve in the survivor of childhood cancer. Pediatric blood & cancer 2009; 53(2): 296-302.
- 29. Thomas-Teinturier C, El Fayech C, Oberlin O, et al. Age at menopause and its influencing factors in a cohort of survivors of childhood cancer: earlier but rarely premature. Human reproduction 2013; 28(2): 488-95.
- 30. Barton SE, Najita JS, Ginsburg ES, et al. Infertility, infertility treatment, and achievement of pregnancy in female survivors of childhood cancer: a report from the Childhood Cancer Survivor Study cohort. The lancet oncology 2013; 14(9): 873-81.
- 31. Reulen RC, Zeegers MP, Wallace WH, et al. Pregnancy outcomes among adult survivors of childhood cancer in the British Childhood Cancer Survivor Study. Cancer epidemiology, biomarkers & prevention : a publication of the American Association for Cancer Research, cosponsored by the American Society of Preventive Oncology 2009; 18(8): 2239-47.
- 32. Edgar AB, Wallace WH. Pregnancy in women who had cancer in childhood. European journal of cancer 2007; 43(13): 1890-4.
- 33. Meirow D, Epstein M, Lewis H, Nugent D, Gosden RG. Administration of cyclophosphamide at different stages of follicular maturation in mice: effects on reproductive performance and fetal malformations. Human reproduction 2001; 16(4): 632-7.
- 34. Duffy CM, Allen SM, Clark MA. Discussions regarding reproductive health for young women with breast cancer undergoing chemotherapy. Journal of clinical oncology : official journal of the American Society of Clinical Oncology 2005; 23(4): 766-73.
- 35. Meirow D, Biederman H, Anderson RA, Wallace WH. Toxicity of chemotherapy and radiation on female reproduction. Clinical obstetrics and gynecology 2010; 53(4): 727-39.
- 36. Ash P. The influence of radiation on fertility in man. The British journal of radiology 1980; 53(628): 271-8.
- 37. Wallace WH, Thomson AB, Kelsey TW. The radiosensitivity of the human oocyte. Human reproduction 2003; 18(1): 117-21.
- 38. Sklar CA, Mertens AC, Mitby P, et al. Premature menopause in survivors of childhood cancer: a report from the childhood cancer survivor study. Journal of the National Cancer Institute 2006; 98(13): 890-6.
- 39. Green DM, Sklar CA, Boice JD, Jr., et al. Ovarian failure and reproductive outcomes after childhood cancer treatment: results from the Childhood Cancer Survivor Study. Journal of clinical oncology : official journal of the American Society of Clinical Oncology 2009; 27(14): 2374-81.



- 40. Green DM, Nolan VG, Kawashima T, et al. Decreased fertility among female childhood cancer survivors who received 22-27 Gy hypothalamic/pituitary irradiation: a report from the Childhood Cancer Survivor Study. Fertility and sterility 2011; 95(6): 1922-7, 7 e1.
- 41. Critchley HO, Wallace WH. Impact of cancer treatment on uterine function. Journal of the National Cancer Institute Monographs 2005; (34): 64-8.
- 42. Signorello LB, Cohen SS, Bosetti C, et al. Female survivors of childhood cancer: preterm birth and low birth weight among their children. Journal of the National Cancer Institute 2006; 98(20): 1453-61.
- 43. Forman EJ, Anders CK, Behera MA. Pilot survey of oncologists regarding treatment-related infertility and fertility preservation in female cancer patients. The Journal of reproductive medicine 2009; 54(4): 203-7.
- 44. Blumenfeld Z, Avivi I, Ritter M, Rowe JM. Preservation of fertility and ovarian function and minimizing chemotherapy-induced gonadotoxicity in young women. Journal of the Society for Gynecologic Investigation 1999; 6(5): 229-39.
- 45. Goodwin T, Elizabeth Oosterhuis B, Kiernan M, Hudson MM, Dahl GV. Attitudes and practices of pediatric oncology providers regarding fertility issues. Pediatric blood & cancer 2007; 48(1): 80-5.
- 46. Quinn GP, Vadaparampil ST, Gwede CK, et al. Discussion of fertility preservation with newly diagnosed patients: oncologists' views. Journal of cancer survivorship : research and practice 2007; 1(2): 146-55.
- 47. Woodruff TK. The Oncofertility Consortium--addressing fertility in young people with cancer. Nature reviews Clinical oncology 2010; 7(8): 466-75.
- 48. Quinn GP, Vadaparampil ST, Bell-Ellison BA, Gwede CK, Albrecht TL. Patient-physician communication barriers regarding fertility preservation among newly diagnosed cancer patients. Social science & medicine 2008; 66(3): 784-9.
- 49. Vadaparampil S, Quinn G, King L, Wilson C, Nieder M. Barriers to fertility preservation among pediatric oncologists. Patient education and counseling 2008; 72(3): 402-10.
- 50. Schover LR, Brey K, Lichtin A, Lipshultz LI, Jeha S. Oncologists' attitudes and practices regarding banking sperm before cancer treatment. Journal of clinical oncology : official journal of the American Society of Clinical Oncology 2002; 20(7): 1890-7.
- 51. Baynosa J, Westphal LM, Madrigrano A, Wapnir I. Timing of breast cancer treatments with oocyte retrieval and embryo cryopreservation. Journal of the American College of Surgeons 2009; 209(5): 603-7.
- 52. Jenninga E, Louwe LA, Peters AA, Nortier JW, Hilders CG. Timing of fertility preservation procedures in a cohort of female patients with cancer. European journal of obstetrics, gynecology, and reproductive biology 2012; 160(2): 170-3.
- 53. Lee S, Ozkavukcu S, Heytens E, Moy F, Oktay K. Value of early referral to fertility preservation in young women with breast cancer. Journal of clinical oncology : official journal of the American Society of Clinical Oncology 2010; 28(31): 4683-6.
- 54. Noyes N, Knopman JM, Melzer K, Fino ME, Friedman B, Westphal LM. Oocyte cryopreservation as a fertility preservation measure for cancer patients. Reproductive biomedicine online 2011; 23(3): 323-33.
- 55. Scott RT, Jr., Elkind-Hirsch KE, Styne-Gross A, Miller KA, Frattarelli JL. The predictive value for in vitro fertility delivery rates is greatly impacted by the method used to select the threshold between normal and elevated basal follicle-stimulating hormone. Fertility and sterility 2008; 89(4): 868-78.
- 56. Riggs RM, Duran EH, Baker MW, et al. Assessment of ovarian reserve with anti-Mullerian hormone: a comparison of the predictive value of anti-Mullerian hormone, follicle-stimulating hormone, inhibin B, and age. American journal of obstetrics and gynecology 2008; 199(2): 202 e1-8.
- 57. Arce JC, La Marca A, Mirner Klein B, Nyboe Andersen A, Fleming R. Antimullerian hormone in gonadotropin releasing-hormone antagonist cycles: prediction of ovarian response and cumulative treatment outcome in good-prognosis patients. Fertility and sterility 2013; 99(6): 1644-53.
- 58. Brodin T, Hadziosmanovic N, Berglund L, Olovsson M, Holte J. Antimullerian hormone levels are strongly associated with live-birth rates after assisted reproduction. The Journal of clinical endocrinology and metabolism 2013; 98(3): 1107-14.



- 59. Brougham MF, Crofton PM, Johnson EJ, Evans N, Anderson RA, Wallace WH. Anti-Mullerian hormone is a marker of gonadotoxicity in pre- and postpubertal girls treated for cancer: a prospective study. The Journal of clinical endocrinology and metabolism 2012; 97(6): 2059-67.
- 60. de Vet A, Laven JS, de Jong FH, Themmen AP, Fauser BC. Antimullerian hormone serum levels: a putative marker for ovarian aging. Fertility and sterility 2002; 77(2): 357-62.
- 61. Freeman EW, Sammel MD, Lin H, Gracia CR. Anti-mullerian hormone as a predictor of time to menopause in late reproductive age women. The Journal of clinical endocrinology and metabolism 2012; 97(5): 1673-80.
- 62. Anders C, Marcom PK, Peterson B, et al. A pilot study of predictive markers of chemotherapy-related amenorrhea among premenopausal women with early stage breast cancer. Cancer investigation 2008; 26(3): 286-95.
- 63. Anderson RA, Cameron DA. Pretreatment serum anti-mullerian hormone predicts long-term ovarian function and bone mass after chemotherapy for early breast cancer. The Journal of clinical endocrinology and metabolism 2011; 96(5): 1336-43.
- 64. Hamre H, Kiserud CE, Ruud E, Thorsby PM, Fossa SD. Gonadal function and parenthood 20 years after treatment for childhood lymphoma: a cross-sectional study. Pediatric blood & cancer 2012; 59(2): 271-7.
- 65. Hagen CP, Vestergaard S, Juul A, et al. Low concentration of circulating antimullerian hormone is not predictive of reduced fecundability in young healthy women: a prospective cohort study. Fertility and sterility 2012; 98(6): 1602-8 e2.
- 66. Chang MY, Chiang CH, Hsieh TT, Soong YK, Hsu KH. Use of the antral follicle count to predict the outcome of assisted reproductive technologies. Fertility and sterility 1998; 69(3): 505-10.
- 67. Wallace WH, Kelsey TW. Ovarian reserve and reproductive age may be determined from measurement of ovarian volume by transvaginal sonography. Human reproduction 2004; 19(7): 1612-7.
- 68. Scheffer GJ, Broekmans FJ, Looman CW, et al. The number of antral follicles in normal women with proven fertility is the best reflection of reproductive age. Human reproduction 2003; 18(4): 700-6.
- 69. Waxman JH, Ahmed R, Smith D, et al. Failure to preserve fertility in patients with Hodgkin's disease. Cancer chemotherapy and pharmacology 1987; 19(2): 159-62.
- 70. Badawy A, Elnashar A, El-Ashry M, Shahat M. Gonadotropin-releasing hormone agonists for prevention of chemotherapy-induced ovarian damage: prospective randomized study. Fertility and sterility 2009; 91(3): 694-7.
- 71. Giuseppe L, Attilio G, Edoardo DN, Loredana G, Cristina L, Vincenzo L. Ovarian function after cancer treatment in young women affected by Hodgkin disease (HD). Hematology 2007; 12(2): 141-7.
- 72. Clowse ME, Behera MA, Anders CK, et al. Ovarian preservation by GnRH agonists during chemotherapy: a metaanalysis. Journal of women's health 2009; 18(3): 311-9.
- 73. Emons G, Grundker C, Gunthert AR, Westphalen S, Kavanagh J, Verschraegen C. GnRH antagonists in the treatment of gynecological and breast cancers. Endocrine-related cancer 2003; 10(2): 291-9.
- 74. Mullen P, Scott WN, Miller WR. Growth inhibition observed following administration of an LHRH agonist to a clonal variant of the MCF-7 breast cancer cell line is accompanied by an accumulation of cells in the G0/G1 phase of the cell cycle. British journal of cancer 1991; 63(6): 930-2.
- 75. D'Angelo A, Amso N. Embryo freezing for preventing Ovarian Hyperstimulation Syndrome. The Cochrane database of systematic reviews 2002; (2): CD002806.
- 76. 2012 Canadian Assisted Reproduction Technology Register (CARTR). 59th Annual Meeting of Canadian Fertility and Andology Society 2013.
- 77. Rudick B, Opper N, Paulson R, Bendikson K, Chung K. The status of oocyte cryopreservation in the United States. Fertility and sterility 2010; 94(7): 2642-6.
- 78. Jain JK, Paulson RJ. Oocyte cryopreservation. Fertility and sterility 2006; 86(4 Suppl): 1037-46.
- 79. Boiso I, Marti M, Santalo J, Ponsa M, Barri PN, Veiga A. A confocal microscopy analysis of the spindle and chromosome configurations of human oocytes cryopreserved at the germinal vesicle and metaphase II stage. Human reproduction 2002; 17(7): 1885-91.
- 80. Cobo A, Rubio C, Gerli S, Ruiz A, Pellicer A, Remohi J. Use of fluorescence in situ hybridization to assess the chromosomal status of embryos obtained from cryopreserved oocytes. Fertility and sterility 2001; 75(2): 354-60.



- 81. Noyes N, Porcu E, Borini A. Over 900 oocyte cryopreservation babies born with no apparent increase in congenital anomalies. Reproductive biomedicine online 2009; 18(6): 769-76.
- 82. Practice Committees of American Society for Reproductive M, Society for Assisted Reproductive T. Mature oocyte cryopreservation: a guideline. Fertility and sterility 2013; 99(1): 37-43.
- 83. Practice Committee of the American Society for Reproductive M, Practice Committee of the Society for Assisted Reproductive T. Ovarian tissue and oocyte cryopreservation. Fertility and sterility 2006; 86(5 Suppl 1): S142-7.
- 84. Kolibianakis EM, Venetis CA, Tarlatzis BC. Cryopreservation of human embryos by vitrification or slow freezing: which one is better? Current opinion in obstetrics & gynecology 2009; 21(3): 270-4.
- 85. Cobo A, Diaz C. Clinical application of oocyte vitrification: a systematic review and meta-analysis of randomized controlled trials. Fertility and sterility 2011; 96(2): 277-85.
- 86. Smith GD, Serafini PC, Fioravanti J, et al. Prospective randomized comparison of human oocyte cryopreservation with slow-rate freezing or vitrification. Fertility and sterility 2010; 94(6): 2088-95.
- 87. Cobo A, Meseguer M, Remohi J, Pellicer A. Use of cryo-banked oocytes in an ovum donation programme: a prospective, randomized, controlled, clinical trial. Human reproduction 2010; 25(9): 2239-46.
- 88. Nagy ZP, Chang CC, Shapiro DB, et al. Clinical evaluation of the efficiency of an oocyte donation program using egg cryo-banking. Fertility and sterility 2009; 92(2): 520-6.
- 89. Al-Inany HG, Youssef MA, Aboulghar M, et al. Gonadotrophin-releasing hormone antagonists for assisted reproductive technology. The Cochrane database of systematic reviews 2011; (5): CD001750.
- 90. Depalo R, Jayakrishan K, Garruti G, et al. GnRH agonist versus GnRH antagonist in in vitro fertilization and embryo transfer (IVF/ET). Reproductive biology and endocrinology : RB&E 2012; 10: 26.
- 91. Fatemi HM, Blockeel C, Devroey P. Ovarian stimulation: today and tomorrow. Current pharmaceutical biotechnology 2012; 13(3): 392-7.
- 92. Pu D, Wu J, Liu J. Comparisons of GnRH antagonist versus GnRH agonist protocol in poor ovarian responders undergoing IVF. Human reproduction 2011; 26(10): 2742-9.
- 93. von Wolff M, Thaler CJ, Frambach T, et al. Ovarian stimulation to cryopreserve fertilized oocytes in cancer patients can be started in the luteal phase. Fertility and sterility 2009; 92(4): 1360-5.
- 94. Cakmak H, Katz A, Cedars MI, Rosen MP. Effective method for emergency fertility preservation: random-start controlled ovarian stimulation. Fertility and sterility 2013.
- 95. Nayak SR, Wakim AN. Random-start gonadotropin-releasing hormone (GnRH) antagonist-treated cycles with GnRH agonist trigger for fertility preservation. Fertility and sterility 2011; 96(1): e51-4.
- 96. Sonmezer M, Turkcuoglu I, Coskun U, Oktay K. Random-start controlled ovarian hyperstimulation for emergency fertility preservation in letrozole cycles. Fertility and sterility 2011; 95(6): 2125 e9-11.
- 97. Buendgen NK, Schultze-Mosgau A, Cordes T, Diedrich K, Griesinger G. Initiation of ovarian stimulation independent of the menstrual cycle: a case-control study. Archives of gynecology and obstetrics 2013; 288(4): 901-4.
- 98. Bedoschi GM, de Albuquerque FO, Ferriani RA, Navarro PA. Ovarian stimulation during the luteal phase for fertility preservation of cancer patients: case reports and review of the literature. Journal of assisted reproduction and genetics 2010; 27(8): 491-4.
- 99. Cakmak H, Rosen MP. Ovarian stimulation in cancer patients. Fertility and sterility 2013; 99(6): 1476-84.
- 100. Oktay K, Turkcuoglu I, Rodriguez-Wallberg KA. GnRH agonist trigger for women with breast cancer undergoing fertility preservation by aromatase inhibitor/FSH stimulation. Reproductive biomedicine online 2010; 20(6): 783-8.
- 101. Humaidan P, Papanikolaou EG, Tarlatzis BC. GnRHa to trigger final oocyte maturation: a time to reconsider. Human reproduction 2009; 24(10): 2389-94.
- 102. Cavagna M, Dzik A. Depot GnRH-agonist trigger for breast-cancer patient undergoing ovarian stimulation re sulted in mature oocytes for cryopreservation: a case report. Reproductive biomedicine online 2011; 22(3): 317-9.
- 103. Humaidan P, Thomsen LH, Alsbjerg B. GnRHa trigger and modified luteal support with one bolus of hCG should be used with caution in extreme responder patients. Human reproduction 2013; 28(9): 2593-4.



- 104. Humaidan P, Polyzos NP, Alsbjerg B, et al. GnRHa trigger and individualized luteal phase hCG support according to ovarian response to stimulation: two prospective randomized controlled multi-centre studies in IVF patients. Human reproduction 2013; 28(9): 2511-21.
- 105. Ortmann O, Weiss JM, Diedrich K. Gonadotrophin-releasing hormone (GnRH) and GnRH agonists: mechanisms of action. Reproductive biomedicine online 2002; 5 Suppl 1: 1-7.
- 106. Prest SJ, May FE, Westley BR. The estrogen-regulated protein, TFF1, stimulates migration of human breast cancer cells. FASEB journal : official publication of the Federation of American Societies for Experimental Biology 2002; 16(6): 592-4.
- 107. Allred CD, Ju YH, Allred KF, Chang J, Helferich WG. Dietary genistin stimulates growth of estrogen-dependent breast cancer tumors similar to that observed with genistein. Carcinogenesis 2001; 22(10): 1667-73.
- 108. Babayan A, Hannemann J, Spotter J, Muller V, Pantel K, Joosse SA. Heterogeneity of estrogen receptor expression in circulating tumor cells from metastatic breast cancer patients. PloS one 2013; 8(9): e75038.
- 109. Gonzalez-Angulo AM, Morales-Vasquez F, Hortobagyi GN. Overview of resistance to systemic therapy in patients with breast cancer. Advances in experimental medicine and biology 2007; 608: 1-22.
- 110. Henderson IC, Garber JE, Breitmeyer JB, Hayes DF, Harris JR. Comprehensive management of disseminated breast cancer. Cancer 1990; 66(6 Suppl): 1439-48.
- 111. Miller WR. Aromatase and the breast: regulation and clinical aspects. Maturitas 2006; 54(4): 335-41.
- 112. Love RR, Van Dinh N, Quy TT, et al. Survival after adjuvant oophorectomy and tamoxifen in operable breast cancer in premenopausal women. Journal of clinical oncology : official journal of the American Society of Clinical Oncology 2008; 26(2): 253-7.
- 113. Howell A, Sims AH, Ong KR, Harvie MN, Evans DG, Clarke RB. Mechanisms of Disease: prediction and prevention of breast cancer-cellular and molecular interactions. Nature clinical practice Oncology 2005; 2(12): 635-46.
- 114. Fisher B, Costantino J, Redmond C, et al. A randomized clinical trial evaluating tamoxifen in the treatment of patients with node-negative breast cancer who have estrogen-receptor-positive tumors. The New England journal of medicine 1989; 320(8): 479-84.
- 115. Goss PE, Ingle JN, Martino S, et al. A randomized trial of letrozole in postmenopausal women after five years of tamoxifen therapy for early-stage breast cancer. The New England journal of medicine 2003; 349(19): 1793-802.
- 116. Howell A, Locker GY. Defining the roles of aromatase inhibitors in the adjuvant treatment of early-stage breast cancer. Clinical breast cancer 2005; 6(4): 302-9.
- 117. Schoolcraft WB, Surrey ES, Minjarez DA, Stevens JM, Gardner DK. Management of poor responders: can outcomes be improved with a novel gonadotropin-releasing hormone antagonist/letrozole protocol? Fertility and sterility 2008; 89(1): 151-6.
- 118. Oktay K, Buyuk E, Davis O, Yermakova I, Veeck L, Rosenwaks Z. Fertility preservation in breast cancer patients: IVF and embryo cryopreservation after ovarian stimulation with tamoxifen. Human reproduction 2003; 18(1): 90-5.
- 119. Checa Vizcaino MA, Corchado AR, Cuadri ME, Comadran MG, Brassesco M, Carreras R. The effects of letrozole on ovarian stimulation for fertility preservation in cancer-affected women. Reproductive biomedicine online 2012; 24(6): 606-10.
- 120. Oktay K, Kan MT, Rosenwaks Z. Recent progress in oocyte and ovarian tissue cryopreservation and transplantation. Current opinion in obstetrics & gynecology 2001; 13(3): 263-8.
- 121. Gosden RG, Baird DT, Wade JC, Webb R. Restoration of fertility to oophorectomized sheep by ovarian autografts stored at -196 degrees C. Human reproduction 1994; 9(4): 597-603.
- 122. Aubard Y. Ovarian tissue xenografting. European journal of obstetrics, gynecology, and reproductive biology 2003; 108(1): 14-8.
- 123. Shaw JM, Bowles J, Koopman P, Wood EC, Trounson AO. Fresh and cryopreserved ovarian tissue samples from donors with lymphoma transmit the cancer to graft recipients. Human reproduction 1996; 11(8): 1668-73.
- 124. Kim SS, Radford J, Harris M, et al. Ovarian tissue harvested from lymphoma patients to preserve fertility may be safe for autotransplantation. Human reproduction 2001; 16(10): 2056-60.



- 125. Foulkes WD, Shuen AY. In brief: BRCA1 and BRCA2. The Journal of pathology 2013; 230(4): 347-9.
- 126. Oktay K, Buyuk E, Rosenwaks Z, Rucinski J. A technique for transplantation of ovarian cortical strips to the forearm. Fertility and sterility 2003; 80(1): 193-8.
- 127. Oktay KH, Yih M. Preliminary experience with orthotopic and heterotopic transplantation of ovarian cortical strips. Seminars in reproductive medicine 2002; 20(1): 63-74.
- 128. Oktay K, Economos K, Kan M, Rucinski J, Veeck L, Rosenwaks Z. Endocrine function and oocyte retrieval after autologous transplantation of ovarian cortical strips to the forearm. JAMA : the journal of the American Medical Association 2001; 286(12): 1490-3.
- 129. Oktay K. Ovarian tissue cryopreservation and transplantation: preliminary findings and implications for cancer patients. Human reproduction update 2001; 7(6): 526-34.
- 130. Oktay K, Karlikaya G. Ovarian function after transplantation of frozen, banked autologous ovarian tissue. The New England journal of medicine 2000; 342(25): 1919.
- 131. Baird DT, Webb R, Campbell BK, Harkness LM, Gosden RG. Long-term ovarian function in sheep after ovariectomy and transplantation of autografts stored at -196 C. Endocrinology 1999; 140(1): 462-71.
- 132. Donnez J, Dolmans MM, Demylle D, et al. Livebirth after orthotopic transplantation of cryopreserved ovarian tissue. Lancet 2004; 364(9443): 1405-10.
- 133. Silber SJ, DeRosa M, Pineda J, et al. A series of monozygotic twins discordant for ovarian failure: ovary transplantation (cortical versus microvascular) and cryopreservation. Human reproduction 2008; 23(7): 1531-7.
- 134. Ernst E, Bergholdt S, Jorgensen JS, Andersen CY. The first woman to give birth to two children following transplantation of frozen/thawed ovarian tissue. Human reproduction 2010; 25(5): 1280-1.
- 135. Sanchez-Serrano M, Crespo J, Mirabet V, et al. Twins born after transplantation of ovarian cortical tissue and oocyte vitrification. Fertility and sterility 2010; 93(1): 268 e11-3.
- 136. Demeestere I, Simon P, Buxant F, et al. Ovarian function and spontaneous pregnancy after combined heterotopic and orthotopic cryopreserved ovarian tissue transplantation in a patient previously treated with bone marrow transplantation: case report. Human reproduction 2006; 21(8): 2010-4.
- 137. Roux C, Amiot C, Agnani G, Aubard Y, Rohrlich PS, Piver P. Live birth after ovarian tissue autograft in a patient with sickle cell disease treated by allogeneic bone marrow transplantation. Fertility and sterility 2010; 93(7): 2413 e15-9.
- 138. Andersen CY, Rosendahl M, Byskov AG, et al. Two successful pregnancies following autotransplantation of frozen/ thawed ovarian tissue. Human reproduction 2008; 23(10): 2266-72.
- 139. Bedaiwy MA, Falcone T. Whole ovary transplantation. Clinical obstetrics and gynecology 2010; 53(4): 797-803.
- 140. Arav A, Natan Y. Directional freezing: a solution to the methodological challenges to preserve large organs. Seminars in reproductive medicine 2009; 27(6): 438-42.
- 141. Courbiere B, Caquant L, Mazoyer C, Franck M, Lornage J, Salle B. Difficulties improving ovarian functional recovery by microvascular transplantation and whole ovary vitrification. Fertility and sterility 2009; 91(6): 2697-706.
- 142. Bromer JG, Patrizio P. Fertility preservation: the rationale for cryopreservation of the whole ovary. Seminars in reproductive medicine 2009; 27(6): 465-71.
- Ives A, Saunders C, Bulsara M, Semmens J. Pregnancy after breast cancer: population based study. Bmj 2007; 334(7586): 194.
- 144. Blakely LJ, Buzdar AU, Lozada JA, et al. Effects of pregnancy after treatment for breast carcinoma on survival and risk of recurrence. Cancer 2004; 100(3): 465-9.
- 145. Helewa M, Levesque P, Provencher D, et al. Breast cancer, pregnancy, and breastfeeding. Journal of obstetrics and gynaecology Canada : JOGC = Journal d'obstetrique et gynecologie du Canada : JOGC 2002; 24(2): 164-80; quiz 81-4.
- 146. Gelber S, Coates AS, Goldhirsch A, et al. Effect of pregnancy on overall survival after the diagnosis of early-stage breast cancer. Journal of clinical oncology : official journal of the American Society of Clinical Oncology 2001; 19(6): 1671-5.



- 147. Han SS, Kim YH, Lee SH, et al. Underuse of ovarian transposition in reproductive-aged cancer patients treated by primary or adjuvant pelvic irradiation. The journal of obstetrics and gynaecology research 2011; 37(7): 825-9.
- 148. Mazonakis M, Damilakis J, Varveris H, Gourtsoyiannis N. Radiation dose to laterally transposed ovaries during external beam radiotherapy for cervical cancer. Acta oncologica 2006; 45(6): 702-7.
- 149. Bisharah M, Tulandi T. Laparoscopic preservation of ovarian function: an underused procedure. American journal of obstetrics and gynecology 2003; 188(2): 367-70.
- 150. Al-Badawi IA, Al-Aker M, AlSubhi J, et al. Laparoscopic ovarian transposition before pelvic irradiation: a Saudi tertiary center experience. International journal of gynecological cancer : official journal of the International Gynecological Cancer Society 2010; 20(6): 1082-6.
- 151. Pahisa J, Martinez-Roman S, Martinez-Zamora MA, et al. Laparoscopic ovarian transposition in patients with early cervical cancer. International journal of gynecological cancer : official journal of the International Gynecological Cancer Society 2008; 18(3): 584-9.
- 152. Morice P, Juncker L, Rey A, El-Hassan J, Haie-Meder C, Castaigne D. Ovarian transposition for patients with cervical carcinoma treated by radiosurgical combination. Fertility and sterility 2000; 74(4): 743-8.
- 153. Nguyen L, Brewer CA, DiSaia PJ. Ovarian metastasis of stage IB1 squamous cell cancer of the cervix after radical parametrectomy and oophoropexy. Gynecologic oncology 1998; 68(2): 198-200.
- 154. Picone O, Aucouturier JS, Louboutin A, Coscas Y, Camus E. Abdominal wall metastasis of a cervical adenocarcinoma at the laparoscopic trocar insertion site after ovarian transposition: case report and review of the literature. Gynecologic oncology 2003; 90(2): 446-9.
- 155. Morris SN, Ryley D. Fertility preservation: nonsurgical and surgical options. Seminars in reproductive medicine 2011; 29(2): 147-54.
- 156. Williams RS, Littell RD, Mendenhall NP. Laparoscopic oophoropexy and ovarian function in the treatment of Hodgkin disease. Cancer 1999; 86(10): 2138-42.
- 157. Ronn R, Holzer HE. Oncofertility in Canada: the impact of cancer on fertility. Current oncology 2013; 20(4): e338-44.
- 158. Ronn R, Holzer HE. Oncofertility in Canada: an overview of Canadian practice and suggested action plan. Current oncology 2013; 20(5): e465-74.